

PII: S0040-4039(97)00676-X

ESR Evidence for a Heterolytic C,O-Bond Cleavage in Models of 4'-DNA Radicals

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Abstract: Photolysis of ketone 6b generates allylic radical 9. The decisive step in this transformation is a heterolytic C,0-bond cleavage from radical 7b that generates radical cation 8 from radical 7b. This ESR study models the anaerobic strand scission of 4'-DNA radicals. © 1997 Elsevier Science Ltd.

The radical induced DNA strand scission by antitumor antibiotics of the bleomycin or enediyne families is achieved by H-atom abstraction from the deoxyribose moiety.¹ Among the deoxyribose radicals the 4'-DNA radical 1 plays an important role since it can cleave the DNA strand even in the absence of O_2 and under neutral conditions.² This spontaneous DNA strand scission is believed to occur via radical cation 2 that reacts with H_2O and generates radical 3.



In spite of the importance of this reaction mechanism there is no ESR spectroscopic evidence for 4'-DNA radical intermediates. Even our attempts to observe ESR spectra using 4'-substituted deoxyribonucleosides 4 and 5 as radical precursors failed.³ However, we have now found that a replacement of the heterocycle b in 4 and 5 by a phenyl group (compound 6) yields a suitable precursor for these ESR experiments.⁴



Figure 2 shows that by photolysis of 6a an ESR spectrum of the modified deoxyribosyl radical 7a can be obtained.⁵ For the assignment of the coupling constants the geometry of radical 7 (R=TMS) was optimized

with the UHF/3-21G^{*} ab initio method, and the Fermi contacts were calculated with the density functionals $BLYP/6-31G^*$ at the unrestricted level of theory.⁶



Fig.1: Calculated conformation of radical 7 (R = TMS).

The small β -coupling constant of 3.6 G (4.7 G calc.) demonstrates the equatorial orientation of the C,Hbond at 3–C. Thus, the C,O-bond at 3–C is in axial position.⁷ The large γ -coupling constant of 8.2 G (7.2 G calcd.) for 1–H can be explained by the axial orientation of the C,H-bond that facilitates the interaction with the spin of the cyclic oxyalkyl radical. Therefore, the phenyl group at 1–C adopts the equatorial position. The quantum chemical calculations also suggest a pyramidalization at the radical carbon center of 7 (Figure 1).⁶



Fig.2: ESR spectrum of 7a (left) measured in benzene at 7°C and the simulation (right) of this spectrum.

A spontaneous, heterolytic cleavage of the C,O-bond at 3-C can occur only with good anionic leaving groups like phosphates.⁸ In order to get more informations about this reaction step, we photolyzed the phosphorylated derivative **6b** in the ESR cavity and observed the formation of the allylic radical **9** (Figure 3).⁵



Coupling Constants (a_H) of 9 in Gauss:

	1–H	2H	3-н	5a,b-H
obs.	32.8	13.3	2.0	8.6
calc.	29.8	14.2	1.8	7.4

The structure of 9 can be unequivocally confirmed by the ESR coupling constants. The small coupling constant of 2.0 G (1.8 G calc.) is typical for a hydrogen at the central carbon of an allylic radical.⁹ The large coupling constant of 32.8 G demonstrates that the C,H-bond at 1–C is in the plane of the allylic π -system so that the phenyl group adopts an equatorial position.



Fig. 3: ESR spectrum of 9 (left) measured in benzene at 7°C and the simulation (right) of this spectrum.

The observation of the allylic radical 9 supports the assumption that radical cation 8 is formed by heterolytic C,O-bond cleavage from radical 7b since is well known that radical cations are acids that deprotonate easily and yield allylic radicals.¹⁰ These ESR spectroscopic measurements were carried out in benzene. Using methanol as solvent the radical cation 8 was trapped by the nucleophilic alcohol.⁴

Conclusion: The ESR spectroscopic measurements using ketone 6 as a precursor for a model of 4'deoxyribonucleoside radicals provide further evidence for a heterolytic strand cleavage of 4'-DNA radicals $(1\rightarrow 2)$ under anaerobic conditions.

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Acknowledgment: This work was supported by the Swiss National Science Foundation. S. P. thanks the Fonds der Chemischen Industrie for a Kekulé-Stipendium.

References and Notes

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(Received in Germany 3 April 1997; accepted 4 April 1997)